Melatonin-Depleted Blood from Premenopausal Women Exposed to Light at Night Stimulates Growth of Human Breast Cancer Xenografts in Nude Rats

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Abstract
The increased breast cancer risk in female night shift workers has been postulated to result from the suppression of pineal melatonin production by exposure to light at night. Exposure of rats bearing rat hepatomas or human breast cancer xenografts to increasing intensities of white fluorescent light during each 12-hour dark phase (0-345 μW/cm²) resulted in a dose-dependent suppression of nocturnal melatonin blood levels and a stimulation of tumor growth and linoleic acid uptake/metabolism to the mitogenic molecule 13-hydroxyoctadecadienoic acid. Venous blood samples were collected from healthy, premenopausal female volunteers during either the daytime, nighttime, or nighttime following 90 minutes of ocular bright, white fluorescent light exposure at 580 μW/cm² (i.e., 2,800 lx). Compared with tumors perfused with daytime-collected melatonin-deficient blood, human breast cancer xenografts and rat hepatomas perfused in situ, with nocturnal, physiologically melatonin-rich blood collected during the night, exhibited markedly suppressed proliferative activity and linoleic acid uptake/metabolism. Tumors perfused with melatonin-deficient blood collected following ocular exposure to light at night exhibited the daytime pattern of high tumor proliferative activity. These results are the first to show that the tumor growth response to exposure to light during darkness is intensity dependent and that the human nocturnal, circadian melatonin signal not only inhibits human breast cancer growth but that this effect is extinguished by short-term ocular exposure to bright, white light at night. These mechanistic studies are the first to provide a rational biological explanation for the increased breast cancer risk in female night shift workers. (Cancer Res 2005; 65(23): 11174-84)

Introduction
The risk of developing breast cancer is up to five times higher in industrialized nations than in underdeveloped countries. Overall, nearly 50% of breast cancers cannot be accounted for by conventional risk factors (1, 2). Westernized nations have increasingly become 24-hour societies with greater numbers of people being exposed to more artificial light during the night both at home and particularly in the workplace (3). Stevens postulated that light exposure at night may represent a unique risk factor for breast cancer in industrialized societies via its ability to suppress the nocturnal production of melatonin by the pineal gland (4, 5). This hypothesis is based on studies showing that melatonin inhibits the development and growth of experimental models of breast cancer, whereas either surgical removal of the pineal gland or exposure to constant light stimulates mammary tumorigenesis in rodents (6). This postulate was further strengthened by the demonstration that physiologic, nocturnal concentrations of melatonin directly inhibited the proliferation of MCF-7 human breast cancer cells in culture (7, 8). Recent epidemiologic studies showing that women working night shifts have a significantly elevated risk of breast cancer presumably due to their potential increased exposure to light at night (9–11) provide additional support for this hypothesis.

Cohen et al. (12) originally hypothesized that diminished pineal function may promote the development of human breast cancer. Their primary argument was that increased pineal calcification, ostensibly leading to lowered melatonin production, was most strongly associated with increased breast cancer risk. Although they were the first to refer to, in general terms, environmental lighting, which necessarily includes both sunlight and artificial light, as one of several endocrine abnormalities that may underlie the development of breast cancer, these workers did not postulate light at night as an etiologic factor. In fact, they incorrectly suggested that altered visual stimulation, as by blindness or darkness, would impair pineal melatonin production leading to unopposed estrogen secretion and increased breast cancer risk. We now know that overall melatonin production is not compromised in blind individuals (13), and breast cancer risk is actually decreased in blind women (14). Subsequent to the Stevens’ hypotheses (4, 5), Kerenyi et al. (15) were the first to propose that “light pollution” may be a potentially important etiologic influence on the genesis of other human cancers. Some indirect support for this postulate was provided by a recent epidemiologic study showing a higher risk of colorectal cancer in women night shift workers (16).

The circadian synthesis and secretion of pineal melatonin during the night results from the rhythmic neuronal activity generated by the central circadian pacemaker in the suprachiasmatic
nuclei of the hypothalamus, which is entrained (i.e., synchronized) by the light/dark cycle. The duration of melatonin production during the night is directly proportional to the length of the dark period (17–19). The alternating light/dark cycle entrains circadian melatonin production to a 24-hour cycle, whereas ocular exposure to light during darkness rapidly suppresses melatonin production depending on the intensity, wavelength, and duration of the light exposure (20, 21). In experimental systems, melatonin plays a fundamental role in the regulation of seasonal reproduction, circadian rhythm activity, retinal physiology, cardiovascular effects, immune function, and cancer (17–19).

It has been generally assumed that the breast cancer–promoting effects of either pinealectomy (i.e., pineal removal) or constant light exposure in rodents are due to the absence of the nocturnal melatonin signal because pharmacologic melatonin replacement either attenuates or prevents these tumorogenic effects (6). Physiologic nocturnal circulating levels of melatonin inhibit the growth of tissue-isolated rat hepatoma 7288CTC via melatonin receptor–mediated suppression of tumor linoleic acid uptake/metabolism to the mitogenic signaling molecule 13-hydroxyoctadecadienoic acid (13-HODE; refs. 22, 23). Pinealectomy or light exposure during darkness prevents these effects (22, 24, 25).

Inasmuch as the Stevens’ hypothesis (4, 5) has not been experimentally tested, we evaluated the response of rats, bearing either rat hepatomas or human breast cancer xenografts, to increasing intensities of ocular light exposure during darkness on tumor growth, linoleic acid uptake/metabolism, and signal transduction activity. More importantly, we also determined the proliferative, metabolic, and signal transduction responses of hepatomas and breast cancer xenografts to perfusion in situ with blood collected from premenopausal human female volunteers whose physiologic circadian melatonin signal was suppressed by ocular exposure to bright, white fluorescent light at night. Our sole rationale for including tissue-isolated rat hepatomas was based on the fact that they exhibit remarkable sensitivity to melatonin, and most of the information we have on the biochemical/molecular mechanisms of melatonin’s anticancer action in vivo has been obtained in this model system (22–25). We wanted the ability to compare the biological/molecular responses to light at night in the new tissue-isolated human breast cancer xenograft model with those in our highly melanolin-sensitive rat liver cancer model serving as a positive control. It is important to note, however, that unlike breast cancer, human liver cancer is much more common in the developing world (i.e., less light pollution) than in industrialized nations (i.e., more light pollution), and its incidence decreases as societies become more westernized (26).

Materials and Methods

Animals, tumors, and histopathology. Adult male Buffalo [BUF(BUF/C0)] rats were implanted with rat hepatocarcinoma syngeneic grafts (Morris 7288CTC), and adult inbred female nude rats (Hsd:RH-rn) were implanted with either steroid receptor–negative (SR−; i.e., no estrogen or progesterone receptor expression) or steroid receptor–positive (SR+) MCF-7 human breast cancer xenografts in a tissue-isolated manner as previously described (27, 28). SR+ tumors evolved over several passages from a subset of SR− xenografts that had become estrogen unresponsive. Both SR− and SR+ human breast cancer xenografts were histopathologically confirmed to be poorly differentiated, grade 3, infiltrating ductal breast adenocarcinomas. Adult male rats (Hsd:Sprague-DawleySD) were used as whole-blood donors for some tumor perfusions. Immunohistochemical staining of cell nuclei for estrogen and progesterone receptor expression in tissue sections of human breast cancer xenografts was done by IMPATH (New York, NY). SR+ tumors were highly estrogen receptor positive (80% staining) and minimally progesterone receptor positive (10% staining); in SR− xenografts no staining for either receptor was evident (data not shown). Animals were maintained in an Association for Assessment and Accreditation of Laboratory Animal Care International–accredited facility on a 12-hour light/12-hour dark cycle (lights on 06:00 to 18:00 hours) and provided with standard laboratory chow and water ad libitum in accordance with an Institutional Animal Care and Use Committee–approved protocol. A series of ceiling fixtures containing 32-W fluorescent tubes (GE Watt Miser, cool white, F40CW.RS.WM) provided 7.5 μW/cm2 of white light at rodent eye level during the light phase.

Tissue-isolated tumor perfusions in situ. SR− MCF-7 human breast cancer xenografts were perfused in situ with rat donor whole blood to which either synthetic melatonin (Sigma, St. Louis, MO), and/or 13-HODE (Cayman Chemicals, Ann Arbor, MI) was added as previously described (22, 23). Sets (three tumors per perfusion) of tissue-isolated rat hepatomas and human breast cancer xenografts were perfused in situ for 1 hour as previously described (22, 23) with whole blood collected from either donor rats or human subjects exposed to each of the lighting conditions outlined below.

Tumor reverse transcription-PCR. Rat hepatoma 7288CTC and SR− human breast cancer xenograft tissues were harvested from rats under nembutal anesthesia between 08:30 and 10:30 hours (zeitgeber time), quickly frozen, and stored at −80°C until subsequent isolation of total RNA. Total cellular RNA was isolated from human breast cancer xenograft tissue and rat hepatoma using the SV Total RNA Isolation System (Promega Corp., Madison, WI). The RNA was quantitated spectrophotometrically, and its integrity was checked by agarose gel. The mRNA was reversed transcribed with Moloney murine leukemia virus reverse transcriptase (Invitrogen Corp., Carlsbad, CA). The resulting cDNA was amplified using the PCR for MT1, or MT2 melatonin receptor product with Taq Polymerase (Promega) at 1.5 mmol/L MgCl2 and a 58°C annealing temperature for 36 cycles using a RoboCycler (Stratagene, La Jolla, CA). Forward and reverse primers used for MT1 and MT2 melatonin receptors in tumor tissues are the same as previously published for rat (29) and for human (30, 31).

Tumor in situ hybridization. At 15:30 hours (zeitgeber time), rat hepatoma and SR− human breast cancer xenograft tissue wedges were immediately harvested from rats under nembutal anesthesia, quickly frozen, and stored at −80°C until subsequent isolation of total RNA. Rat hepatoma and SR− human breast cancer tissue wedges were embedded in ornithine carbamyl transferase compound, rapidly frozen in 2-methylbutane kept in dry ice, and stored frozen at −80°C until tissue sections were cut and thaw mounted onto glass slides. Tumor sections were processed for in situ hybridization using digoxigenin-labeled oligonucleotide antisense and sense probes for MT1 and MT2 melatonin receptors as previously described (32). The MT1 oligonucleotide (GGGGCTGCTACTGGA-GAGTTCCCCGTTTGCGAG, mer 31) corresponds to bases 108 to 138 of the partial rat MT1 receptor sequence (Genbank accession no. U14409) and to bases 518 to 548 of the human MT1 receptor sequence (Genbank accession no. U14108). The MT2 oligonucleotide (CGGGTCATATTTCTGAGA-GACCCACAAAGAAA, mer 31) corresponds to bases 111 to 141 of the partial rat MT2 receptor sequence (Genbank accession no. U28218) and to bases 537 to 556 of the human MT2 receptor sequence (Genbank accession no. U23431).

Nocturnal light exposure studies in rats and effects on tumor growth. Groups of tumor-bearing rats were housed in light exposure chambers that contained two separate solid-state, electromagnetic fluorescent ballasts with rapid-start, cool-white lamps connected to separate 24-hour timers and separated by a metal baffle (20). One ballast/lamp system (GE Watt Miser, F43CW-RS-WM, 34-W bulb) in each chamber was set up to provide a direct, steady, bright light stimulus (345 μW/cm2) at the animals’ eye level during the light portion of a 12-hour light/12-hour dark cycle. The second ballast/lamp system (GE Starcoat, F32T8-SP-11, 32-W bulb) was adjustable with a combination of neutral filter density filter material (CINELG 3403, N-9) and electronic dimmer modules to emit steady, indirect light reflected off of the rear chamber wall.
measured at the animals' eye level during the dark phase. The entire chamber was painted with Pro-Mar flat white paint (Sherwin Williams, St. Simon Island, GA), which has a high content of titanium dioxide that reflects light relatively uniformly across the visible spectrum. The light intensities during each 12-hour dark phase were different in each chamber as follows: 0 (total darkness), 0.02, 0.05, 0.06, 0.08, and 345 μW/cm (constant light). These intensities were measured and regularly monitored at animals' eye level in the center of each cage using a Model IL 1400a Radiometer/Photometer (International Light, Inc., Newburyport, MA). Animals were exposed to the various light intensities beginning 2 weeks before tumor implantation and continuing thereafter until the end of each tumor growth period. Throughout each experiment, animal cage positions were rotated on a daily basis to minimize the potential effects of slight variations in the target, reflected light intensity within each chamber.

Nocturnal light exposure studies in human subjects and effects on tumor proliferative activity. At Thomas Jefferson University (Philadelphia, PA), healthy, premenopausal female volunteers (n = 12; mean age, 22.6 ± 0.6 years) were recruited without regard to either race or ethnic background and signed approved institutional review board consent forms. All subjects were free of any medication, including birth control pills, melatonin supplements, and/or β-blockers, and had regular sleep patterns. Each subject had antecubital venous blood samples drawn by a phlebotomist during the daytime (10:00 to 14:00 hours), nighttime (02:00 to 04:00 hours) following 2 hours of complete darkness, and nighttime (03:30 hours) following 2 hours of complete darkness, and nighttime (05:30 hours) following 90 minutes of ocular, bright, white light exposure at 580 μW/cm (i.e., 2,800 lx) at eye level reflected from the wall. This exposure protocol is described in detail elsewhere (33). On the day following the final blood collection, whole-blood samples were shipped on ice via overnight express mail to the Bassett Research Institute (Cooperstown, NY), where they were stored refrigerated with constant gentle vortexing until tumor perfusions were done 24 hours later.

Assay of blood hormone levels in rats and human subjects. Rat serum and human plasma melatonin levels were measured via RIA as previously described (21, 23). Rat serum corticosterone and estradiol levels were measured using a commercial RIA kits (Diagnostic Products Corp., Los Angeles, CA).

Determination of tumor growth, linoleic acid uptake, and 13-hydroxoyctadecadienoic acid formation. Tumor growth rates, uptake of linoleic acid, and formation of 13-HODE were determined as previously described (21, 23). Due to variations in the absolute circulating concentrations of linoleic acid, tumor linoleic acid uptake is expressed as a percent of the actual arterial supply of linoleic acid to the tumor.

Determination of tumor cyclic AMP levels, DNA content, and [3H]thymidine incorporation into DNA. Tumor levels of cyclic AMP (cAMP) were determined via ELISA (Amersham Biosciences, Piscataway, NJ) and [3H]thymidine incorporation into DNA, and DNA content were determined as previously described (22, 23, 28).

Determination of tumor mitogen-activated protein/extracellular signal-regulated kinase signal-regulated kinase and extracellular signal-regulated kinase 1/2 activation. Western blot analysis of tumor mitogen-activated protein/extracellular signal-regulated kinase kinase (MEK) and extracellular signal-regulated kinase 1/2 (ERK1/2) activation was done as previously described (34).

Statistical analyses. Statistical differences among group means were determined by a one-way ANOVA followed by Student-Neuman-Keuls post hoc test. Differences in the slopes of regression lines (i.e., tumor growth rates) among groups were determined by regression analyses and tests for parallelism (Student’s t test). Four-variable parabolic fluence (dose)-response curve fitting of the above variables was done as previously described (35). Differences were considered statistically significant at P < 0.05.

Results

Melatonin responsiveness of human breast cancer xenografts perfused in situ. We previously reported that tissue-isolated rat hepatoma 7288CTC was highly responsive to the inhibitory effects of physiologic nocturnal melatonin concentrations with respect to tumor linoleic acid uptake/metabolism and proliferative activity following perfusion in situ (22, 23). Furthermore, the tumor inhibitory effects of melatonin were blocked by both MT1 and MT2 receptor antagonists (36). We now show that SR’-MCF-7 human breast cancer xenografts were also very sensitive to the inhibitory actions of nocturnal melatonin concentrations by perfusing tissue-isolated tumors in situ with exogenous synthetic melatonin (1 nmol/L; see perfusion diagram in Fig. 5B). Following 2 hours of perfusion with melatonin, tumor cAMP levels were reduced by 40%, whereas both tumor linoleic acid uptake and 13-HODE formation were completely blocked (Table 1). These changes were accompanied by a decrease in ERK1/2 activation (Fig. 1) and a 71% decrease in tumor [3H]thymidine incorporation into DNA, which were completely prevented by the coperfusion of tumors with both melatonin and 13-HODE (Fig. 1; Table 1). These findings indicated that, similar to rat hepatoma 7288CTC (22, 23), SR’- MCF-7 human breast cancer xenografts were highly sensitive to a nocturnal, physiologic concentration of melatonin, which is consistent with the expression of MT1 receptors in SR’-MCF-7 cells (37). This high degree of melatonin sensitivity was also exhibited in tissue-isolated SR’-MCF-7 breast cancer xenografts that had evolved from and retained the histopathologic phenotype of the SR’-MCF-7 parental tumor (see Materials and Methods) and also expressed MT1 receptors (see below). Therefore, SR’-MCF-7 xenografts were used in testing the effects of melatonin suppression, via ocular light exposure during darkness in either rats or premenopausal human subjects, on breast cancer growth progression.

Melatonin receptor mRNA expression in rat hepatoma and human breast cancer xenografts. Although human cancer cell lines, including SR’-MCF-7 breast cancer cells (37), have been shown to express either the MT1 or MT2 mammalian melatonin receptors (6), information about the expression of either of these melatonin receptors in solid tumors in vivo is extremely limited (38). This prompted us to determine with both reverse transcription-PCR and in situ hybridization techniques whether one or both of these melatonin receptors are expressed in tissue-isolated rat hepatoma 7288CTC and SR’ and SR’-MCF-7 human breast cancer xenografts. PCR products of the expected size were obtained using primers specific for MT1 and MT2 melatonin receptors for both rat hepatoma and human breast cancer xenograft tissue (SR’, Fig. 2A). No product was observed from a reaction with RNA alone. Hybridization signal for antisense MT1-specific oligonucleotide was found in both rat hepatoma and SR’-MCF-7 human breast cancer xenograft tissue, whereas only rat hepatoma showed specific hybridization signal with MT2 antisense specific oligo-probe (Fig. 2B). These hybridization signals were localized primarily to tumor epithelial cells. From a functional standpoint, however, melatonin receptor antagonist studies reveal that whereas rat hepatoma expresses both MT1 and MT2 melatonin receptors (39), human breast cancer xenografts only express functional MT1 melatonin receptors (40).

Melatonin suppressive and tumor stimulatory responses to different intensities of nocturnal light exposure in rat hepatoma. We next determined the fluence-response (i.e., dose-response) effects of light exposure during darkness on signal transduction activity, linoleic acid uptake/metabolism, and growth in tissue-isolated rat hepatoma 7288CTC in view of its exquisite melatonin sensitivity (22–24). Groups of male rats housed in specially designed light exposure chambers (Fig. 3A) were exposed to different intensities of light during the dark phase of an
alternating 12-hour light/12-hour dark cycle beginning 2 weeks before tumor implantation and continuing thereafter until the end of the tumor growth experiment. Following 2 weeks of exposure of rats to increasing intensities of reflected light (0 [total darkness], 0.02, 0.05, 0.06, 0.08, and 345 [constant bright light] μW/cm²) measured at rodent eye level during darkness, there was a dose-dependent increase in the percent suppression of serum melatonin levels (Fig. 4A). Continued exposure of rats to these light intensities following tumor implantation resulted in a corresponding dose-related increase in tumor growth rates (0.62-1.55 g/d; Fig. 3B and Fig. 4E). [3H]thymidine incorporation into DNA (Fig. 4C), and DNA content (data not shown). In addition, the time to tumor onset decreased as the light intensity increased (Fig. 3B). Similarly, tumor linoleic acid uptake and 13-HODE formation (Fig. 4B and D), ERK1/2 activation (Fig. 3B), and cAMP levels (Table 2) increased as a function of increased light intensities during the dark phase. Four-variable parabolic fluence-response curves (Fig. 4A-E) showed a tight correlation (r² = 0.93-0.99) between percent melatonin suppression and tumor linoleic acid uptake, 13-HODE production, [3H]thymidine incorporation into DNA, and growth rates. The ED₅₀ values (i.e., half maximal suppression or stimulation) for melatonin suppression and tumor stimulatory responses to light during darkness ranged from 0.06 to 0.09 μW/cm². No significant effects of light exposure at any intensity was observed on serum corticosterone levels during the mid-dark period, albeit levels were depressed in the constant light-exposed group (Table 3). These results confirmed, in a highly melanotin-sensitive murine tumor model (22–24), that tumor signal transduction, linoleic acid uptake, and 13-HODE formation and growth could be modulated in a dose-dependent manner by changes in light intensity exposure during the dark phase of a diurnal lighting cycle.

Melatonin suppressive and tumor stimulatory responses to different intensities of nocturnal light exposure in human breast cancer xenografts. Previously, we reported that the female nude rat exhibits a robust nocturnal circadian melatonin rhythm, similar to that in humans in terms of its amplitude, phasing, and duration, that is completely suppressed by constant bright light (i.e., 300 lx or ~120 μW/cm²) at rodent eye level (ref. 27). Furthermore, in nude rats bearing tissue-isolated SR⁺ MCF-7 human breast cancer xenografts, constant bright light exposure resulted in a significant increase in tumor linoleic acid uptake/metabolism to 13-HODE and a marked stimulation of tumor growth compared with control rats on 12-hour light/12-hour dark cycle (27). We found that like tissue-isolated SR⁺ MCF-7 xenografts (Fig. 1; Table 1), SR⁺ xenografts (Fig. 5E and G) perfused in situ were highly responsive to the tumor-inhibiting effects of a physiologic, nocturnal circulating concentration of melatonin (see below). We assessed the dose-response effects of light exposure during darkness on growth, linoleic acid uptake/metabolism to 13-HODE, and signal transduction activity in SR⁺ breast cancer xenografts under the same lighting conditions as outlined above for rats bearing rat hepatoma 7288CCT. As the light intensity increased, there was a corresponding increase in the percent suppression of serum melatonin levels in nude rats (Fig. 4F). There was also an accompanying marked, dose-related increase in tumor growth rates (0.22-0.74 g/d; Fig. 3C and Fig. 4J), [3H]thymidine incorporation into DNA (Fig. 4H), and DNA content (data not shown). Latency to tumor onset decreased as the intensity of light during darkness increased (Fig. 3C). Tumor linoleic acid uptake and 13-HODE formation (Fig. 4G and I), ERK1/2 activation (Fig. 3C), and cAMP levels (Table 2) increased as a function of the light intensity exposure during the dark phase. As in the case of rat hepatoma, four-variable parabolic fluence-response curves (Fig. 4F-J) showed a tight correlation (r² = 0.97-0.99) between percent melatonin suppression and tumor linoleic acid uptake, 13-HODE formation, and the percent suppression of serum melatonin levels in nude rats (Fig. 4F).

### Table 1. Response of SR⁺ MCF-7 human breast cancer xenografts to perfusion in situ with a physiologic nocturnal concentration of melatonin

<table>
<thead>
<tr>
<th>Treatment</th>
<th>[3H]Thymidine incorporation (dpm/μg DNA)</th>
<th>Linoleic acid uptake (% supply)</th>
<th>13-HODE production (ng/min/g)</th>
<th>cAMP (nmol/g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls</td>
<td>47.4 ± 3.9</td>
<td>16.7 ± 1.7</td>
<td>0.97 ± 0.17</td>
<td>0.55 ± 0.11</td>
</tr>
<tr>
<td>Melatonin</td>
<td>13.6 ± 1.6</td>
<td>0</td>
<td>0</td>
<td>0.33 ± 0.12*</td>
</tr>
<tr>
<td>Melatonin + 13-HODE</td>
<td>7.4 ± 0.6</td>
<td>0</td>
<td>333.17 ± 19.02</td>
<td>0.68 ± 0.06</td>
</tr>
<tr>
<td>13-HODE</td>
<td>7.8 ± 0.6</td>
<td>17.2 ± 3.2</td>
<td>363.18 ± 10.62</td>
<td>0.78 ± 0.17</td>
</tr>
</tbody>
</table>

NOTE: Effects of perfusion of tissue-isolated SR⁺ MCF-7 human breast cancer xenografts in situ with synthetic melatonin (1 nmol/L) in the presence or absence of 13-HODE (12 μg/mL) on [3H]thymidine incorporation into DNA, linoleic acid uptake, 13-HODE formation, and cAMP levels. Individual tumors (n = 3-4) were perfused over a 2-hour period (06:30 to 08:30 h) with whole blood collected from tumor-free donor rats during the early light phase when endogenous melatonin levels were low. Synthetic melatonin and 13-HODE were added to the whole-blood perfusate to achieve final concentrations of 1 nmol/L (232 pg/mL) and 12 μg/mL, respectively. Values are means ± SE.

*P < 0.05 vs controls and melatonin + 13-HODE (comparisons of relevance).

**Figure 1.** Tumor expression of ERK1/2. Western blot analysis of the effects of perfusion of tissue-isolated SR⁺ MCF-7 human breast cancer xenografts in situ for 1 hour with melatonin (1 nmol/L) in the presence or absence of 13-HODE (12 μg/mL) on ERK1/2 (mitogen-activated protein kinase p44/p42) activation. Top, phosphorylated ERK1/2; bottom, total ERK1/2. Total protein (27 μg) was loaded per well. Controls (lanes 1-3), melatonin-treated (lanes 4-6), melatonin + 13-HODE (lanes 7-9), 13-HODE (lanes 10-12), and MW markers (lane 13). See Table 2 for further details.
[3H]thymidine incorporation into DNA, and growth rates. ED_{50} values for these processes ranged from 0.06 to 0.07 A^2/W/cm^2. No significant effects of light exposure at any intensity was observed on serum either corticosterone or estradiol levels during the mid-dark period, albeit levels of these hormones tended to be decreased or increased, respectively, in the constant light-exposed group (Table 3). These results confirmed that suppression of physiologic, nocturnal melatonin levels was dependent upon the magnitude of the light intensity to which nude rats were exposed during the dark phase. This suppression of melatonin levels translated to an intensity-dependent stimulation of human breast cancer signal transduction activity, linoleic acid uptake/metabolism, and growth.

Proliferative activity in rat hepatoma and human breast cancer xenografts perfused in situ with blood collected from human subjects following ocular exposure to bright white light at night. We next evaluated the acute effect of the human circadian melatonin signal and its suppression by light at night on signal transduction activity, linoleic acid uptake/metabolism, and proliferative activity in rat hepatomas and human breast cancer xenografts. This involved the unique approach of perfusing these tumors, in situ, with whole blood collected from healthy, premenopausal human volunteers either during the daytime, nighttime, or following exposure to 580 μW/cm^2 (i.e., 2,800 lx) of white fluorescent light at night (ref. 33; Fig. 5A and B). As expected, plasma melatonin titers in human volunteers were 5-fold higher during the night compared with daytime levels of melatonin (Fig. 5C). The perfusion of tissue-isolated rat hepatomas and human breast cancer xenografts in situ with melatonin-rich blood collected during the night resulted in tumor cAMP levels that were reduced by 86% compared with control tumors perfused with melatonin-deficient blood collected during the daytime (Fig. 5D and E). Similarly, tumor linoleic acid uptake and 13-HODE formation were reduced by as much as 96% following perfusion with melatonin-rich blood collected from human subjects during darkness compared with control tumors perfused with daytime-collected, melatonin-deficient blood (Fig. 5D-G). Furthermore, tumor incorporation of [3H]thymidine into DNA following perfusion with melatonin-rich blood collected during the night was decreased by 63% to 73% compared with control tumors perfused with daytime-collected, melatonin-deficient blood (Fig. 5D-G). In contrast, tumors perfused with melatonin-deficient blood obtained from subjects during the night following 90 minutes of light exposure evinced signal transduction activity,
linoleic acid uptake/metabolism, and proliferative activity that remained at the same levels as control tumors perfused with daytime-collected, melatonin-deficient blood (Fig. 5D–G). Similar results were obtained in a small subset of SR+ human breast cancer xenografts perfused in situ with blood collected from human subjects during the night and following exposure to light at night (data not shown).

We then determined whether the tumor-suppressive effects of nighttime-collected, melatonin-rich blood from human subjects were, in fact, due to melatonin. Tissue-isolated tumors were perfused with melatonin-deficient blood, collected at night following 90 minutes of bright light exposure, to which a nocturnal concentration of synthetic melatonin was added (i.e., 500 pmol/L or 116 pg/mL). The tumor-stimulating effects of melatonin-deficient blood collected at night following light exposure were prevented with this melatonin replacement strategy (Fig. 5D–G). Moreover, the addition of the nonselective MT1/MT2 melatonin receptor antagonist S20928 (36) to the blood perfusate collected during the night completely blocked the tumor-suppressive effects of this nocturnal, physiologically melatonin-rich blood (Fig. 5D–G).

Discussion
Our results show that tissue-isolated rat hepatoma 7288CTC expressed mRNA transcripts for both MT1 and MT2 melatonin receptors both of which are functionally expressed and mediate appropriate tumor responses to nocturnal, physiologic melatonin concentrations and to melatonin receptor antagonists (22, 23, 39). Tissue-isolated SR+ MCF-7 human breast cancer xenografts also expressed mRNA transcripts for both MT1 and MT2 melatonin receptors, whereas SR- xenografts expressed only MT1 melatonin receptors that are functionally expressed and mediate appropriate tumor responses to melatonin and melatonin receptor antagonists (40). In the present investigation, we confirmed that rat hepatoma 7288CTC, and both SR+ and SR- human breast cancer xenografts were highly sensitive to the ability of nocturnal, physiologic circulating levels of melatonin to suppress tumor proliferative activity in vivo. Additionally, ocular exposure of tumor-bearing rats to increasing intensities of light during darkness resulted in a dose-dependent suppression of melatonin blood levels that translated to a light intensity–dependent stimulation of signal transduction activity and linoleic acid–dependent rat liver and human breast tumor growth rates. Rat hepatomas and human breast cancer xenografts perfused in situ with nocturnal, physiologically melatonin-rich blood collected from human, premenopausal female volunteers during the night exhibited suppressed signal transduction and linoleic acid–dependent proliferative activity. Suppressed tumor growth signaling activity was absent in tumors perfused with melatonin-deficient blood collected from human subjects during the daytime or following ocular exposure to white light at night.

The circadian system exerts an important influence over many physiologic and metabolic activities, such as the sleep/wake cycle, body temperature, intermediary metabolism, and endocrine functions, and a number of disease processes, including myocardial infarction, stroke, and asthma (19, 41). Indirect evidence is emerging that disruption of circadian clock activity is associated with cancer in humans, and that desynchronization of internal timekeeping adversely tips the host/cancer balance in favor of tumor development and growth in experimental models of tumorigenesis (42, 43). Cancer cells themselves express the same clock genes found in the suprachiasmatic nuclei (44), and many common molecular elements are shared between the circadian clock and cell cycle (45, 46). A mutation in the circadian clock gene per2 results in an increase in spontaneous tumorigenesis in mice (46), whereas an elevated risk of premenopausal breast cancer in females results in an increase in spontaneous tumorigenesis in mice (46). Additionally, ocular exposure of tumor-bearing rats to light during darkness.

![Figure 3. Ocular exposure of tumor-bearing rats to light during darkness. A, schematic representation of a side, cut-a-way view of a light exposure chamber showing the relationship of the two separate solid-state, electromagnetic fluorescent ballasts with rapid-start, cool-white fluorescent lamps separated by a metal baffle. The ballast/lamp system adjacent to the hinged front wall of the chamber provides a direct, steady, bright light stimulus (345 μW/cm²) at the animals’ eye level during the light portion of a 12-hour light/12-hour dark cycle. The second ballast/lamp system adjacent to the rear wall was adjustable with neutral density filters and electronic dimmer modules to emit steady, indirect light reflected off of the rear chamber wall measured at the animals’ eye level during the dark phase only. Families of tumor growth curves reflecting the effects of exposure of rats bearing either (B) hepatoma 7288CTC (male rats) or (C) SR+ MCF-7 human breast cancer xenografts (nude female rats) to increasing intensities of light during the entire dark phase (12 hours) of a 12-hour light/12-hour dark cycle on tumor growth and ERK1/2 activation (phosphorylation). Light exposures began 2 weeks before tumor implantation and continued throughout each tumor growth period in the light chambers. At the end of each growth period for light treatment groups (dark, 0.05 μW/cm² and 345 μW/cm²), tumors were freeze-clamped under liquid N₂ and ERK1/2 activation was determined by Western blotting (one representative tumor from each of these groups is depicted). Points, mean (n = 6 rats, tumors per group); bars, SE. Slopes of regression lines and tests for parallelism among growth curves show that rats exposed to 345, 0.08, 0.06, and 0.05 μW/cm² had significantly (P < 0.05) faster tumor growth rates compared with those exposed to 0.02 and 0.00 μW/cm² (dark control). All intensities of light versus dark control and each other (P < 0.05).](www.aacrjournals.org)
cancer has been associated with length polymorphism in the clock gene per3 (47). In spite of this progress, however, no definitive outputs linking the human circadian clock with the processes governing human malignancy, particularly breast cancer, have been identified. Moreover, the way in which circadian disruption by light at night may lead to an increased risk of breast cancer, as suggested by the increased breast cancer risk in night shift workers (9–11), is unknown.

The pineal synthesis and secretion of melatonin represents an important nocturnal output component of the central circadian

Figure 4. Four-variable parabolic curves. Relationship between increasing intensities of white fluorescent light and suppression of plasma melatonin levels in either male rats (A) or female nude rats (F) following 2 weeks of light exposure during the entire dark phase (12 hours) of a 12-hour light/12-hour dark cycle. Plasma samples were collected via decapitation when melatonin peaks at the mid-dark phase (2400 hours). The fluence-response relationship between light intensity (µW/cm²) and percent change (suppression) in melatonin levels compared with the dark control levels was determined by the fluence-response curve formula: Y = [A₁ - A₂] / [1 + (X/X₅₀)²] + A₂, where A₁ = theoretical initial Y value for curve; A₂ = theoretical final Y value for curve; ED₅₀ = X value of curve midpoint Y value; P = power function describing curve slope; R² = correlation coefficient. Points, means (n = 6); bars, SE. Effects of light exposure on male rats bearing hepatoma 7988CTC (B-E) or female nude rats bearing SR/C0MCF-7 human breast cancer xenografts (G-J) to increasing intensities of light during the entire dark phase (12 hours) of a 12-hour light/12-hour dark cycle on tumor linoleic acid metabolism and growth activity. Each of the four-variable parabolic curves shows the fluence-response relationship between light intensity (µW/cm²) and tumor linoleic acid uptake (% supply), 13-HODE formation (ng/min/g), [3H]thymidine incorporation into DNA (dpm/µg DNA; determined at the end of each tumor growth period in Fig. 3), and growth rates (g/d; slopes of the regression lines of tumor growth curves of Fig. 3) as determined by the formula above. Points, mean (n = 6); bars, SE.
pacemaker mechanism (17–19). Melatonin exerts an oncostatic action on a variety of neoplasms in experimental models of cancer in vitro and in vivo, most notably in those involving mammary carcinogenesis (6–8, 37, 48). Based on human breast cancer perfusion studies described here, however, we now present the first evidence establishing the fact that physiologic, nocturnal melatonin concentrations inhibit cancer proliferative activity in vivo, irrespective of tumor type and/or steroid receptor status.

As discussed above, the development and/or growth of experimental mammary or liver cancer is accelerated in response to either pinealectomy or ocular exposure to constant bright light, ostensibly due to the lack of a nocturnal melatonin signal (25). In both rodents and humans, light exposure suppresses pineal melatonin production in an intensity-, wavelength-, and duration-dependent manner (20, 21, 52). The present results represent the first demonstration that the rates of both rat hepatoma and human breast cancer growth increase as a function of a light intensity-dependent suppression of nocturnal circulating melatonin levels. The highly correlated ED_{50} values among the fluence-response curves indicate a remarkable degree of fidelity between light during darkness-induced melatonin suppression and the stimulation of signal transduction activity, linoleic acid uptake/metabolism, and tumor growth. In the case of human breast cancer xenografts, only a 15% to 25% (0.040–0.055 μW/cm²) suppression of the circadian amplitude of nocturnal melatonin was required to cause the first significant stimulation of growth and linoleic acid uptake/metabolism to 13-HODE. The lack of significant light-induced changes in either plasma corticosterone or estradiol levels during darkness indicated that the tumor growth results were not due to light during darkness-induced alterations in the levels of adrenocortical or ovarian hormones known to influence breast cancer risk (53, 54). The lack of a significant effect of light during darkness on estradiol levels, and the fact that both rat hepatomas and SR+ breast cancer xenografts responded to light-induced nocturnal melatonin suppression, convincingly argues against the postulate that increased breast cancer risk, resulting from pineal hypofunction, and reduced melatonin output, is due to hyperestrogenism (12). It is important to note that Stevens (5) subsequently modified his original hypothesis (4), involving light-induced nocturnal melatonin suppression and hyperestrogenism, to include an effect of light at night in suppressing the direct antiproliferative effects of physiologic melatonin on human breast cancer growth (7, 8).

### Table 2. Tumor levels of total cAMP in response to increasing light intensities during darkness

<table>
<thead>
<tr>
<th>Light intensity (μW/cm²)</th>
<th>Tumor cAMP concentration (nmol/g)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Rat hepatoma 7288CTC</td>
</tr>
<tr>
<td>Dark</td>
<td>0.18 ± 0.01</td>
</tr>
<tr>
<td>0.02</td>
<td>0.45 ± 0.06</td>
</tr>
<tr>
<td>0.05</td>
<td>0.50 ± 0.06</td>
</tr>
<tr>
<td>0.06</td>
<td>0.53 ± 0.03</td>
</tr>
<tr>
<td>0.08</td>
<td>0.48 ± 0.03</td>
</tr>
<tr>
<td>Constant light, 345</td>
<td>0.72 ± 0.04</td>
</tr>
</tbody>
</table>

NOTE: Effects of exposure of rats bearing either hepatoma 7288CTC (male rats) or SR- MCF-7 human breast cancer xenografts (nude female rats) to increasing intensities of light during the entire dark phase (12 hours) of a 12-hour light/12-hour dark cycle on tumor cAMP concentrations. At the end of the growth period for each of the light treatment groups, tumors were freeze-clamped under liquid N₂ and then assayed for cAMP via an ELISA. See Figs. 2 and 3 legends for further details. Value are means ± SE; n = 6 rats (tumors per group).

*P < 0.05, hepatoma vs dark control.

1P < 0.05, breast vs dark control (except 0.08 μW/cm²).

1P < 0.05, hepatoma vs all other groups.

1P < 0.05, breast vs all other groups (except 0.08 μW/cm²).

### Table 3. Serum levels of corticosterone and estradiol in rats exposed to increasing light intensities during darkness

<table>
<thead>
<tr>
<th>Light intensity (μW/cm²)</th>
<th>Nude rat serum corticosterone (ng/mL)</th>
<th>Nude rat serum estradiol (pg/mL)</th>
<th>Buffalo rat serum corticosterone (ng/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Constant light, 345</td>
<td>196.2 ± 53.5</td>
<td>25.1 ± 11.3</td>
<td>53.25 ± 23.7</td>
</tr>
<tr>
<td>0.08</td>
<td>265.0 ± 28.17</td>
<td>19.9 ± 10.1</td>
<td>120.83 ± 19.6</td>
</tr>
<tr>
<td>0.06</td>
<td>322.7 ± 43.8</td>
<td>27.1 ± 16.8</td>
<td>105.08 ± 25.8</td>
</tr>
<tr>
<td>0.05</td>
<td>289.3 ± 41.8</td>
<td>20.3 ± 10.78</td>
<td>155.5 ± 19.1</td>
</tr>
<tr>
<td>0.02</td>
<td>266.8 ± 34.7</td>
<td>25.1 ± 5.9</td>
<td>138.75 ± 26.8</td>
</tr>
<tr>
<td>Dark</td>
<td>266.1 ± 34.4</td>
<td>19.3 ± 10.7</td>
<td>111.08 ± 29.5</td>
</tr>
</tbody>
</table>

NOTE: See Figs. 3 and 4 legends for further details. Blood samples were taken during the middle of the true dark or subjective dark phase. Mean serum hormone (±SE); n = 6 rats per group. There were no significant differences in hormone levels among any of the groups exposed to different light intensities.
It would be impossible, both logistically and ethically, to test the light at night/breast cancer hypothesis in humans by chronically exposing healthy women to bright light each night under controlled laboratory conditions. Thus, the perfusion of tissue-isolated tumors in situ with whole blood collected from healthy human female volunteers represents a unique approach to directly assess the effect of the human nocturnal, circadian melatonin output signal on proliferative, metabolic, and signal transduction.
activity in human breast cancer xenografts. In fact, in our opinion, this is the only strategy available to evaluate the potential consequences of circadian disruption of the human melanatonin signal, by ocular exposure of human subjects to bright, white light at night, on human tumor growth activity. Consistent with the results of perfusion in situ of rat hepatomas (22, 23) and human breast cancer xenografts (see above) with rat donor whole blood to which exogenous melanatonin was added at a nocturnal physiologic concentration, the present findings show that physiologically melatonin-rich blood, collected during the night from human subjects, markedly inhibited signal transduction activity (i.e., cAMP, MEK, ERK1/2), linoleic acid uptake/metabolism to 13-HODE and proliferative activity (i.e., [3H]hymidine incorporation into DNA, DNA content) in both rat hepatoma and human breast cancer xenografts (i.e., both SR and SR+ tumors). Each of these oncostatic effects were completely eliminated upon suppression of the nocturnal, circadian melanatonin signal following the exposure of human subjects to 90 minutes of 2,800 lx (580 µW/cm²) of indirect, white fluorescent light. These tumor-suppressive effects of nighttime-collected, melatonin-rich blood were also totally blocked by the nonselective melanatonin receptor (MT₁/MT₂) antagonist S20928 (36), indicating that they were, in fact, due to the presence of physiologically elevated levels of circulating melanatonin acting via a melanatonin receptor-mediated process. This is further supported by the ability of melatonin, added at a physiologic nocturnal concentration, to melatonin-deficient blood collected following light exposure at night, to restore the tumor-inhibitory responses.

To our knowledge, the present findings are the first to establish a role for the nocturnal, physiologic melanatonin signal from the pineal in the prevention and progression of an overt human disease. More specifically, melatonin is now the first soluble, nocturnal anticancer signal to be identified in humans that directly links the central circadian clock with some of the important mechanisms regulating human breast carcinogenesis and possibly the progression of other malignancies as well. These findings also provide the first definitive nexus between the exposure of healthy premenopausal female human subjects to bright white light at night and the enhancement of human breast oncogenesis via disruption (i.e., suppression) of the circadian, oncosstatic melanatonin signal. The suppression of circadian melanatonin production by ocular exposure to bright white light at night, leading to augmented nocturnal tumor uptake of dietary linoleic acid and its conversion to mitogenically active 13-HODE, can now be afforded serious consideration as a new risk factor for human breast cancer (4, 5) and a significant public health issue (55). The high nocturnal dietary intake of fat, particularly linoleic acid, reported for night shift workers (56, 57), coupled with melatonin suppression by exposure to light at night provide a firm mechanistic basis upon which to explain, in part, the increased risk of breast cancer in some women who work night shifts for many years (9–11). Thus, strategies to preserve the integrity of the circadian melanatonin signal (i.e., avoidance of bright light at night, intelligent lighting design, circadian-timed physiologic melanatonin supplementation) coupled with modifications in nocturnal dietary fat intake may offer a unique approach to the prevention of breast cancer, and perhaps other melanatonin-sensitive cancers, in our increasingly 24-hour society.

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References

melatonin receptor-mediated inhibition of tumor
linoleic acid metabolism to the growth signaling
molecule 13-hydroxyoctadecadienoic acid and the
potential role of phytomelanin. Carcinogenesis 2004;
24. Dauchy RT, Black DE, Dauer LA, Brainard GC,
Krause JA. Dim light during darkness stimulates tumor
progression by enhancing tumor fatty acid uptake and
25. Black DE, Brainard GC, McGowan T, Kesseling J.
In: relationship between light and development and
growth of internal solid cancers: a review of current
research and the potential implications for lighting
practice. EPBI 101:1162. Palo Alto (CA) and Conyers
27. Black DE, Dauchy RT, Sauer LA, Krause JA, Brainard
GC. Growth and fatty acid metabolism of human breast
cancer (MCF-7) xenografts in nude rats: impact of
constant light-induced nocturnal melatonin suppres-
28. Sauer LA, Dauchy RT, Identification of linoleic
and arachidonic acids as factors in hyperlipemic blood
that increase [H]thymidine incorporation in hepato-
ma 7288CTC perfused in situ. Cancer Res 1988;48:
3106–11.
29. Zalatan F, Krause JA, Black DE. Inhibition of
isoproterenol-induced lipolysis in rat inguinal adipoc-
etes in vitro by physiological melatonin via a
receptor-mediated mechanism. Endocrinology 2001;
142:3783–90.
30. Reppert M, Weaver DR, Ebisawa T. Cloning and
characterization of a mammalian melatonin receptor
that mediates reproductive and circadian responses.
31. Reppert M, Codson C, Mahle C, Weaver DR,
Slaugenhaupt SA, Guasela JF. Molecular characterization
of a second melatonin receptor expressed in human
retina and brain: the Mel1b melatonin receptor. Proc
32. Hunt AE, Al-Ghoul WM, Gillette MU, Dubocovich
ML. Activation of MT2 melatonin receptors in the rat
suprachiasmatic nucleus phase advances the circadian
Biological effects of architectural lighting and their
associated energy utilization. J Illumin Engineer Soc
34. Sauer LA, Dauchy RT, Black DE, et al. Conjugated
linoleic acid isomers and trans fatty acids inhibit fatty
acid transport in hepatoma 7288CTC and inguinal fat
spectrum for melatonin regulation in humans: evidence
for a novel circadian photoreceptor. J Neurosci 2001;21:
6405–12.
New selective ligands of human cloned melatonin
MT1 and MT2 receptors. Arch Pharmc (Weinheim)
2003;367:553–61.
37. Ram PT, Dai J, Yuan L, et al. Involvement of the mt1
melatonin receptor in human breast cancer. Cancer Lett
2002;179:141–50.
expression of high-affinity melatonin receptors (MT1)
in normal and malignant breast tissue. Am J Clin
39. Dauchy RT, Black DE, Sauer LA, Krause JA,
Davidson J.K. Evidence for MT2 and MT2 receptor-
mediated pathway in melatonin inhibition of fatty
acid transport by hepatoma 7288CTC perfused in vitro
supplementation in rats and a human subject sup-
presses the growth activity of steroid receptor negative
human breast cancer xenografts in female nude rats via
an MT1 receptor-mediated suppression of signal trans-
duction and linoleic acid uptake and metabolism [abstract).
41. Hrushesky WJM. Timing is everything. The Sciences
42. Hrushesky WJM. The temporal organization of life:
the impact of multifrequency non-linear biologic time
structure upon the host-cancer balance. Jpn J Clin Oncol
43. Rutter J, Reick M, McKnight SL. Metabolism and
the control of circadian rhythms. Annu Rev Biochem
44. Fu L, Lee CC. The circadian clock: pacemaker and
45. Canaple L, Kakizawa T, Laudeit V. The days and
Period 3 structural variation: a circadian biomarker
associated with breast cancer in young women. Cancer
Epidemiol Biomarkers Prev 2005;14:268–70.
47. Hill SM, Barn PT, Molis TM, Spriggs LW. Melatonin
at the neuronal and astroglial level. In: Shafii M, Shafi SL,
editors. Melatonin in psychiatric and neoplastic dis-
acid metabolite, (13β-hydroxyoctadecadienoic acid:
its metabolism, and its possible role in cancer. J Nutr
2001;131:2500S–G.
49. Reppert M, Weaver DR, Ebisawa T. Cloning and
determination of a second mammalian melatonin
receptor that mediates reproductive and circadian responses.
50. Reppert M, Codson C, Mahle C, Weaver DR,
Slaugenhaupt SA, Guasela JF. Molecular characterization
of a second melatonin receptor expressed in human
retina and brain: the Mel1b melatonin receptor. Proc
51. Hunt AE, Al-Ghoul WM, Gillette MU, Dubocovich
ML. Activation of MT2 melatonin receptors in the rat
suprachiasmatic nucleus phase advances the circadian
52. Brainard GC, Richardson BA, Petterborg LJ, Reiter
RJ. The effect of different light intensities on pineal
53. Sephton SE, Sapolsky RM, Kraemer HC, Spiegel D.
Diurnal cortisol rhythm as a predictor of breast cancer
54. Clemons M, Goss P. Estrogen and the risk of breast
55. Pauley SM. Lighting for the human circadian
clock: recent research indicates that lighting has
come to be a public health issue. Med Hypotheses 2004;
64:1–52.
responses to nocturnal eating in men are affected by
57. Morgan L, Hampton S, Gibbs M, Arendt J. Circadian
aspects of postprandial metabolism. Chronobiol Int